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# A Reexamination of Species Boundaries between Solanum megistacrolobum and S. toralapanum (Solanum sect. Petota, series Megistacroloba): Morphological Data

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ABSTRACT. Solanum megistacrolobum and S. toralapanum are two phenetically similar wild potato (Solanum sect. Petota) species, classified in series Megistacroloba, that together are distributed from southern Peru to northwestern Argentina. They have variously been synonymized, recognized as varieties of S. megistacrolobum, or recognized as distinct species. This study examines 26 morphological characters of 115 living accessions of these two species, plus six accessions of two related species in series Megistacroloba, S. boliviense and S. sogarandinum. All measurements were taken from living accessions planted in a common garden plot. Solanum megistacrolobum and S. toralapanum could be distinguished phenetically only by a combination of the 21 characters that showed statistically significant differences between the two taxa. No individual characters provided consistent discrimination, and there was frequent overlap of "species-specific" characters between taxa. Our results most closely fit the contemporary treatment of S. megistacrolobum and S. toralapanum at the varietal level.

The genus Solanum L. is estimated to contain 1000–1100 species (D'Arcy 1991), including the potato and its wild relatives (Solanum sect. Petota Dumort.). The latest taxonomic interpretation of sect. Petota (Hawkes 1990) includes 232 species divided into 21 taxonomic series, but there are widely conflicting treatments of sect. Petota. They differ regarding species boundaries, the rank of infraspecific taxa, the placement of species into series, and hypotheses of interspecific hybridization (Spooner and Sytsma 1992; Spooner and van den Berg 1992a).

One series containing species illustrating many of these alternative hypotheses is series Megistacroloba Cárd. and Hawkes, distributed from southern Peru through Bolivia to northwestern Argentina. The series is generally characterized by plants with a rosette habit, or with short stems, terminal leaf lobes larger than the laterals and with lateral leaf lobes typically broadly decurrent on the basiscopic side, long pedicels with articulation near the apex, and sub-stellate to rotate, usually purple corollas. All species are diploid (2n = 24), self-incompatible, and largely interfertile (Buck 1966; Hawkes 1990; Ochoa 1990; Pandey 1960). Correll (1962) includes 11 species in series Megistacroloba, Gorbatenko (1989) 16 species, Hawkes (1990) 11 species, and Ochoa (1990) seven species and one nomenclaturally designated hybrid species. One taxonomic controversy within series *Megistacroloba* concerns the taxonomic status and hypotheses of hybridization of *S. megistacrolobum* Bitter and *S. toralapanum* Cárd. and Hawkes. Ochoa (1984) recognizes them as conspecific, but later (Ochoa 1990) regards them as varieties of a single species [as *S. megistacrolobum* var. *megistacrolobum* and var. *toralapanum* (Cárd. and Hawkes) Ochoa]. They have been recognized as distinct species by Correll (1962), Gorbatenko (1989) and Hawkes (1990).

Solanum megistacrolobum is characterized by a rosette habit or with stems slightly elongate; leaves simple to pinnately lobed with 0-10 decurrent lateral leaflets, paired or unpaired; peduncles 0-5.5 cm long; pedicels 2-9 cm long; inflorescence with 1-8 flowers; calyces with upright posture and corollas lilac to purple, pentagonal to sub-stellate (Correll 1962; Hawkes 1990; Hawkes and Hjerting 1989; Ochoa 1990). Also, the leaves of *S. megistacrolobum* have a weak to strong "parsley-like" odor (Hawkes 1990; Hawkes and Hjerting 1989; Ochoa 1990). There is much variation, however, for all of these features. Solanum megistacrolobum has been recognized as a potential source of valuable traits such as frost resistance and drought tolerance (Brücher 1959; Hawkes and Hjerting 1989; Ochoa 1990).

Solanum toralapanum is morphologically sim-

TABLE 1. Morphological characters used by prior taxonomists to distinguish S. megistacrolobum and S. toralapanum.

Character	S. megistacrolobum	S. toralapanum	
1. Primary lateral leaflet	Narrowly decurrent	Broadly decurrent	
2. Length of peduncle	0-5.5 cm	6-8 cm	
3. Posture of calyx acumen	Straight to appressed	Spreading to recurved	
4. "Parsley-like" odor of leaf	Strong	None	

ilar to S. megistacrolobum. It is characterized by a rosette habit or with stems slightly elongate; leaves simple to pinnately lobed with 0-8 decurrent lateral leaflets, paired or unpaired; peduncles 6-8 cm long; pedicels 1-5 cm long; inflorescence with 1-several (exact number not stated) flowers; calyces with spreading to recurved posture and corollas lilac to purple, rotate-pentagonal to sub-stellate (Correll 1962; Hawkes 1990; Hawkes and Hjerting 1989; Ochoa 1990). The leaves of S. toralapanum lack the parsley-like odor characteristic of S. megistacrolobum. There is much variation for all of these features within S. toralapanum except for the parsley-like leaf odor, which is not known to occur in this taxon.

The main characters that have been used in the past to distinguish S. megistacrolobum from S. toralapanum (Hawkes 1990; Hawkes and Hjerting 1989; Ochoa 1990) are outlined in Table 1. Our preliminary observations of living germplasm accessions at the National Research Support Program-6 (NRSP-6; formerly known as the Inter-Regional Potato Introduction Project, IR-1) however, noted much inter-and intraspecific morphological variability. There also is some distributional and possible ecological differentiation between the two species. Solanum toralapanum grows from 3000-4500 m, often more to the east of S. megistacrolobum on the east-facing Andean mountain slopes and valleys (Fig. 1; Table 2). Our accessions of S. toralapanum are from areas 3-10, 16, 17, 19, 21, 26, and 33 (Fig. 1). Both species are mapped from regions 4, 7, 16, 19, 21, 26, and 33. Solanum megistacrolobum grows from 2700-4450 m, on upland valleys and intermontane basins in the eastern Andean cordillera (Hawkes and Hjerting 1989; Fig. 1). Hawkes and Hjerting (1989) state that S. toralapanum grows in wetter sites than S. megistacrolobum, but Ochoa (1990) lists the habitat of S. megistacrolobum as moist.

Brücher (1959) notes extensive morphologi-

cal variability within S. megistacrolobum that he ascribes to different growth responses in different environments. Correll (1962) recognizes an apparent morphological connection between S. megistacrolobum and S. toralapanum but felt the extremes should be maintained as separate species. These morphological and distributional patterns led Ochoa (1984) to synonymize S. toralapanum under S. megistacrolobum. Johns et al. (1987) invalidly treat S. toralapanum as a variety of S. megistacrolobum. Hawkes and Hjerting (1989) state that plants phenotypically intermediate between S. megistacrolobum and S. toralapanum breed true and are probably the result of introgression between them. Ochoa (1990) cites unpublished studies at the International Potato Center, Peru (CIP), documenting extensive intraspecific variability within S. megistacrolobum and S. toralapanum that he attributes to a combination of genotypic variability and environmental plasticity. Ochoa (1990) formally transferred S. toralapanum to a variety of S. megistacrolobum.

There are many hypotheses about the interrelationships and hybridization of S. megistacrolobum and S. toralapanum. None of these distinguish phenetic from cladistic concepts. Brücher (1959) hypothesizes hybridization between S. megistacrolobum and S. acaule Bitter (2n = 48; series Acaulia). Correll (1962) states that S. megistacrolobum and S. toralapanum are closely allied. He also hypothesizes occasional hybridization between the two, as well as hybridization of S. megistacrolobum with S. acaule and S. sanctae-rosae Hawkes (2n = 24; series Megistacroloba). Hawkes and Hjerting (1969) hypothesize hybridization or introgression between *S*. megistacrolobum and S. boliviense Dunal (2n = 24; series Megistacroloba), S. acaule, S. infundibuliforme Philippi (2n = 24; series Cuneoalata), S.microdontum Bitter (2n = 24; series Tuberosa) and S. gourlayi Hawkes (2n = 24; series Tuberosa) or S. oplocense Hawkes [with diploid (2n = 24), tet-

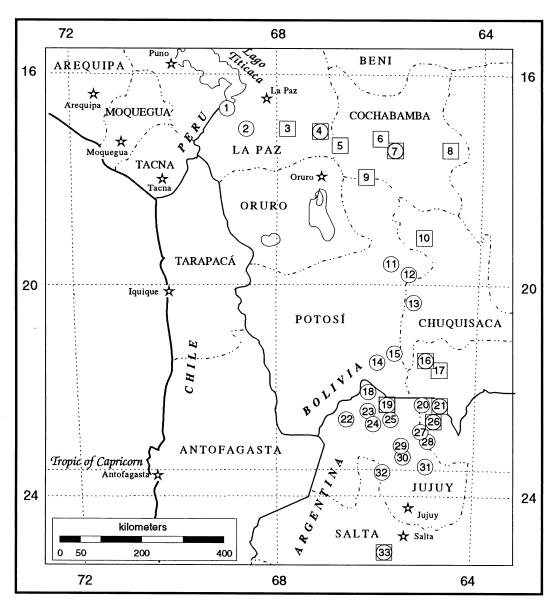


FIG. 1. Map showing the 33 generalized areas of the accessions of Solanum acaule subsp. aemulans, S. boliviense, S. megistacrolobum and S. toralapanum examined in this study or in a companion study (Giannattasio and Spooner 1994) using nuclear RFLPs (see Table 2). Some species other than S. megistacrolobum and S. toralapanum were collected outside of this map area. Examined accessions of S. megistacrolobum are from circled areas; S. toralapanum are from squared areas; accessions of both species were examined from areas with an overlayed circle and square.

raploid (2n = 48), and hexaploid (2n = 72) cytotypes; series *Tuberosa*]. They designate *S.* × *brucheri* Correll (2n = 24; series *Acaulia*) as a natural hybrid between *S. megistacrolobum* and *S. acaule.* Ugent (1970) hypothesizes that *S. raphanifolium* Cárd. and Hawkes (2n = 24; series

Megistacroloba) is of hybrid origin between S. canasense Hawkes (2n = 24; series Tuberosa) and S. megistacrolobum. Solanum megistacrolobum is hypothesized to be a diploid parent (with S. stenotomum Juz. and Buk.; 2n = 24; series Tuberosa) of the cultivated diploid species, S. ajanhuiri Juz.

TABLE 2. Accessions examined. Those examined in this morphological study are marked with an M in the second column. Those marked with a D were used in a separate DNA study (Giannattasio and Spooner 1994). Vouchers are deposited at the herbarium of the National Research Support Program, Sturgeon, Bay, Wisconsin.  $^1$  acl = S. acaule, blb = S. bulbocastanum, blv = S. boliviense, brd = S. brevidens, can = S. canasense, meg = S. megistacrolobum, rap = S. raphanifolium, sgr = S. sogarandinum, tor = S. toralapanum.  $^2$  M = Accession used in the morphological study. D = Accession used in the DNA study.  $^3$  United States Department of Agriculture Plant Introduction Numbers.  $^4$  Generalized map areas. See Fig. 1.  $^5$  Collector unknown; accession from the Erin Baur Sortiment Genebank, Germany.  $^6$  Collector unknown; accession from the Commonwealth Potato Collection, Scotland.

Taxon <sup>1</sup>	Study <sup>2</sup>	PI <sup>3</sup>	Area <sup>4</sup>	Collector	Locality
1. meg	M, D	473361	1	Hawkes et al. 5058	Bolivia. La Paz: 31 km from Desaguadero, on road to Puno; 3950 m; 16°34'S, 69°02'W.
2. meg	M, D	473360	2	Hawkes et al. 4853	Bolivia. La Paz: 4 km past Caquiaviri on road to Río Desaguadero; 4000 m; 17°03'S, 68°38'W.
3. meg	M, D	498258	4	Ochoa 11916	Bolivia. La Paz: between Tres Cruces and Río Seco; 3500 m; 17°06′S, 67°21′W.
4. meg	M, D	498263	7	Ochoa 12098	Bolivia. Cochabamba: Toralapa; 3600 m; 17°26'S, 65°43'W.
5. meg	M	265873	11	EBS <sup>5</sup> 1793	Bolivia. Potosí: Potosí; 19°35'S, 65°45'W.
6. meg	M	265874	11	EBS 1808	Bolivia. Potosí: 8 km from Potosí on road to Don Diego; 19°35′S, 65°45′W.
7. meg	M, D	473356	11	Hawkes et al. 4317	Bolivia. Potosí: below Potosí on road to Lecherías; 3950 m; 19°35'S, 65°45'W.
8. meg	M	473358	11	Hawkes et al. 4589	Bolivia. Potosí: 0.5 km below Potosí on road to Lecherías; 3950 m; 19°35'S, 65°45'W.
9. meg	M	473359	11	Hawkes et al. 4590	Bolivia. Potosí: 0.7 km below Potosí on road to Lecherías; 3950 m; 19°35'S, 65°45'W.
10. meg	M, D	210034	12	Hjerting 1028	Bolivia. Potosí: Alkatayo; 19°50'S, 65°30'W.
11. meg	M	283082	13	EBS 1787	Bolivia. Chuquisaca: Sivingamayo; 3800 m; 20°20'S, 65°04'W.
12. meg	M	473357	13	Hawkes et al. 4323	Bolivia. Chuquisaca: 126 km on road from Potosí to Carmargo, near Payacota; 3500 m; 20°24'S, 65°18'W.
13. meg	M	473497	13	Astley 49	Bolivia. Chuquisaca: 130 km from Potosí on road to Camargo; 19°35'S, 65°45'W.
14. meg	M, D	498259	14	Ochoa 11960	Bolivia. Potosí: Sud Chichas; 4250 m, 20°40'S, 67°00'W.
15. meg	M	498260	15	Ochoa 11962	Bolivia. Potosí: Sud Chichas; pass of Machu Cruz in route to Tupiza; 4100 m; 21°27'S, 65°43'W.
16. meg	M	498261	15	Ochoa 11963	Bolivia. Potosí: Sud Chichas; pass of Machu Cruz in route to Tupiza; 4100 m; 21°27'S, 65°43'W.
17. meg	M	498262	15	Ochoa 11964	Bolivia. Potosí: Sud Chichas; pass of Machu Cruz in route to Tupiza; 4100 m; 21°27'S, 65°43'W.
18. meg	M	265578	16	Correll 654	Bolivia. Tarija: Iscayachi; 21°31'S, 65°03'W.
19. meg	M, D	545897	16	Hoopes et al. 99	Bolivia. Tarija: Méndez; 4 km from Iscayachi on road to Tarija; 3500 m; 21°27'S, 64°58'W.
20. meg	M, D	545898	16	Hoopes et al. 101	Bolivia. Tarija: Aviles; 11 km from Iscayachi on road to Villazón; 3540 m; 21°34′S, 65°02′W.
21. meg	M	458347	18	Hoffman 1682	Argentina. Jujuy: 19 km E of Santa Catalina on the way to El Angosto; 4200 m; 21°55′S; 66°08′W.

TABLE 2. Continued.

Taxon <sup>1</sup>	Study <sup>2</sup>	PI <sup>3</sup>	Area <sup>4</sup>	Collector	Locality
22. meg	M	458349	18	Okada 3933	Argentina. Jujuy: 24 km E of Santa Catalina; 4200 m; 21°57′S, 66°04′W.
23. meg	M	473116	18	Okada 3914	Argentina. Jujuy: 10 km W of Santa Catalina; 4000 m; 21°57′S, 66°04′W.
24. meg	M	473117	18	Okada 3917	Argentina. Jujuy: 10 km W of Santa Catalina; 4000 m; 21°57'S, 66°04'W.
25. meg	M, D	473118	18	Okada 3926	Argentina. Jujuy: 10 km W of Santa Catalina; 4000 m; 21°57'S, 66°04'W.
26. meg	M	473119	18	Okada 3931	Argentina. Jujuy: 24 km W of Santa Catalina; 4200 m; 21°57'S, 66°04'W.
27. meg	M	473120	18	Okada 3934	Argentina. Jujuy: 24 km W of Santa Catalina; 4200 m; 21°57'S, 66°04'W.
28. meg	M	473122	18	Okada 3983	Argentina. Jujuy: Oratorio; 4000 m; 22°05'S, 66°07'W.
29. meg	M, D	473121	18	Okada 3977	Argentina. Jujuy: Cabrerillas W to Oratorio; 3900 m; 22°05′S, 66°07′W.
30. meg	M, D	458348	19	Okad <b>a</b> 3906	Argentina. Jujuy: Tafna; 3600 m; 22°06′S, 65°44′W.
31. meg	M	473115	19	Okada 3886	Argentina. Jujuy: Río Chico; 3620 m; 22°08'S, 65°44'W.
32. meg	M	473135	19	Okada 4442	Argentina. Jujuy: Quebrada del Chorro; 3900 m, 22°08'S, 65°38'W.
33. meg	M	473136	19	Okada 4442	Argentina. Jujuy: on slope from ravine of Chorro; 3900 m; 22°08'S, 65°38'W.
34. meg	M, D	473137	19	Okada <b>4444</b>	Argentina. Jujuy: top of Quebrada del Chorro, along path from Casa Colorada to Molulo; 3700 m, 22°08'S, 65°38'W.
35. meg	M, D	275148	20	Hjerting 255	Argentina. Jujuy: between Lizoite and Abra de Lizoite; 3600 m; 22°15'S, 65°13'W.
36. meg	M	473112	20	Hoffman 1663	Argentina. Jujuy: Cajas; 4000 m; 22°15'S, 65°18'W.
37. meg	M, D	435072	20	Okada 5823	Argentina. Salta: Santa Victoria; 3800 m; 22°15'S, 64°58'W.
38. meg	M, D	473158	21	Okada 5839	Argentina. Salta: on the way from Lizoite; 3400 m; 22°16'S, 65°11'W.
39. meg	M	473139	22	Okada 4452	Argentina. Jujuy: Pucará; 3700 m; 22°24'S, 66°34'W.
40. meg	M	473153	23	Okada 5772	Argentina. Jujuy: 3 km SW of Timon Cruz; 4140 m; 22°13'S, 66°09'W.
41. meg	M	473154	23	Okada 5774	Argentina. Jujuy: 1 km W of Oros; 4000 m; 22°16'S, 66°21'W.
42. meg	M	473155	23	Okada 5775	Argentina. Jujuy: 3 km W of Oros; 3980 m; 22°16'S, 66°21'W.
43. meg	M	473156	23	Okada 5776	Argentina. Jujuy: 4 km W of Oros; 3940 m; 22°16'S, 66°21'W.
44. meg	M	473157	23	Okada 5782	Argentina. Jujuy: 4 km W of Oros; 3840 m; 22°16'S, 66°21'W.
45. meg	M	458350	24	Okada 3989	Argentina. Jujuy: Rinconada; 3800 m; 22°26'S, 66°10'W.
46. meg	M	458351	24	Okada 4018	Argentina. Jujuy: Rinconada, near Abra de Fundiciones; 4200 m; 22°33′S, 66°17′W
47. meg	M	473123	24	Okada 3992	Argentina. Jujuy: Sierra de Rinconada; 3900 m; 22°26'S, 66°07'W.

TABLE 2. Continued.

Taxon <sup>1</sup>	Study <sup>2</sup>	PI <sup>3</sup>	Area <sup>4</sup>	Collector	Locality
48. meg	M	473124	24	Okada 4006	Argentina. Jujuy: Rinconada; 4000 m; 22°26'S, 66°10'W.
49. meg	M	473125	24	Okada 4008	Argentina. Jujuy: Rinconada; 4000 m; 22°26'S, 66°10'W.
50. meg	M	473126	24	Okada 4024	Argentina. Jujuy: 8 km SW of Rinconada; 4020 m; 22°26'S, 66°10'W.
51. meg	M	473152	24	Okada 5526	Argentina. Jujuy: Salveayoc, 5 km SW of Rinconada; 4000 m; 22°26'S, 66°10'W.
52. meg	M	473162	24	Okada 5996	Argentina. Jujuy: on the way from Orosmayo; 4100 m; 22°35'S, 66°21'W.
53. meg	M, D	458346	25	Hoffman 1629	Argentina. Jujuy: Cochinoca; 3650 m; 22°44'S, 65°54'W.
54. meg	M	458352	25	Okada 4421	Argentina. Jujuy: El Angosto; 4000 m; 24°02'S, 65°35'W.
55. meg	М	473131	25	Okada 4422	Argentina. Jujuy: Laguna River, on the way from Casa Colorado to El Durazno; 4000 m; 22°28'S, 65°41'W.
56. meg	M	473160	25	Okada 5956	Argentina. Jujuy: Sierra de Cochinoca; 3700 m; 22°34'S, 65°53'W.
57. meg	M	473161	25	Okada 5957	Argentina. Jujuy: Sierra de Cochinoca; 3700 m; 22°34'S, 65°53'W.
58. meg	M	473114	26	Hoffman 1729	Argentina. Salta: Poscaya; 3420 m; 22°28'S, 65°06'W.
59. meg	M	265879	27	EBS 1783	Argentina. Jujuy: Tres Cruces; 3800 m; 22°55'S, 65°35'W.
60. meg	M	275147	27	Hjerting 366	Argentina. Jujuy: between Condor and Abra del Condor; 3800 m; 22°53′S, 65°16′W.
61. meg	D	233125	27	CPC <sup>6</sup> 2482	Argentina. Jujuy: between Condor and Abra del Condor; 3800 m; 22°53′S, 65°16′W.
62. meg	M, D	473109	27	Hoffman 1598	Argentina. Jujuy: Cuesta Azul Pampa; 3400 m; 23°04'S, 65°24'W.
63. meg	M	473111	27	Hoffman 1618	Argentina. Jujuy: Condor; 4000 m; 22°54'S, 65°17'W.
64. meg	M	473113	27	Hoffman 1701	Argentina. Jujuy: Condor; 4000 m; 22°54'S, 65°17'W.
65. meg	M	473127	27	Okada 4358	Argentina. Jujuy: near La Cueva; 3900 m; 22°55'S, 65°22'W.
66. meg	M	473132	27	Okada 4428	Argentina. Jujuy: La Cueva; 3900 m; 22°55'S, 65°22'W.
67. meg	M	473133	27	Okada 4430	Argentina. Jujuy: La Cueva; 3900 m; 22°55'S, 65°22'W.
68. meg	M	473134	27	Okada 4431	Argentina. Jujuy: La Cueva; 3900 m; 22°55'S, 65°22'W.
69. meg	M	473148	27	Okada 4567	Argentina. Jujuy: between Esquina Blanca and Tres Cruces; 3700 m; 22°57′S, 65°32′W.
70. meg	M	473150	27	Okada 4573	Argentina. Jujuy: 14 km NE of Iturbe; 3700 m; 22°59'S, 65°21'W.
71. meg	M	473524	27	EBS 2945	Argentina. Jujuy: valley of Humahuaca, Azul Pampa; 23°04'S, 65°24'W.
72. meg	М	473149	27	Okada <b>4</b> 569	Argentina. Jujuy: 3 km past Abra de Azul Pampa towards Iturbe; 3700 m; 22°58'S, 65°26'W.
73. meg	M	473140	28	Okada 4460	Argentina. Jujuy: Palca de Aparso; 3800 m; 23°06′S, 65°06′W

TABLE 2. Continued.

Taxon <sup>1</sup>	Study <sup>2</sup>	PI <sup>3</sup>	Area <sup>4</sup>	Collector	Locality
74. meg	M	473163	28	Okada 6734	Argentina. Jujuy: Aparso; 3800 m; 23°06'S, 65°10'W.
75. meg	M	473164	28	Okada 6737	Argentina. Jujuy: 2 km S of Aparso; 3650 m; 23°06'S, 65°10'W.
76. meg	M	473110	29	Hoffman 1615	Argentina. Jujuy: Chalcamayo; 3880 m; 23°14'S, 65°33'W.
77. meg	M	473141	30	Okada 4506	Argentina. Jujuy: Sierra Alta; 3660 m; 23°23'S, 65°32'W.
78. meg	M	473142	30	Okada 4509	Argentina. Jujuy: Sierra Alta; 3200 m; 23°23'S, 65°32'W.
79. meg	M	473143	30	Okada 4512	Argentina. Jujuy: Sierra Alta; 3300 m; 23°23'S, 65°32'W.
80. meg	M	473144	30	Okada 4514	Argentina. Jujuy: Sierra Alta; 3300 m; 23°23'S, 65°32'W.
81. meg	M	473145	30	Okada 4515	Argentina. Jujuy: Sierra Alta; 3300 m; 23°23'S, 65°32'W.
82. meg	M	473147	30	Okada 4524	Argentina. Jujuy: Sierra Alta; 3300 m; 23°23'S, 65°32'W.
83. meg	M, D	473146	30	Okada 4520	Argentina. Jujuy: Sierra Alta; 3300 m; 23°23'S, 65°32'W.
84meg	M, D	473128	31	⊙kada 4362	Argentina. Jujuy: along path from Casa Colora da de Molulo, Ventura River; 4000 m; 23°33'S, 65°12'W.
85. meg	M	473129	31	Okada 4411	Argentina. Jujuy: Piedra Parada; 3700 m; 23°35′S, 65°10′W.
86. meg	M, D	500031	31	Okada 7606	Argentina. Jujuy: 7 hr E of Tilcara by mule, between Peña Parada and Portrero de Ven- tura; 3500 m; 23°35'S, 65°13'W.
87. meg	M	473130	32	Okada 4415	Argentina. Jujuy: on the path from El Duraz- no to Casa Colorada; 3800 m; 23°10'S, 65°39'W.
88. meg	M, D	473159	32	Okada 5954	Argentina. Jujuy: 21 km from Abra Pampa along Route 7; 3700 m; 23°35′S, 65°51′W.
89. meg	M, D	275149	33	Hjerting 316	Argentina. Jujuy: Piedra Molino; 3700 m; 25°10'S, 65°52'W.
90. meg	M	500029	33	Okada 7504	Argentina. Salta: Cuesta del Obispo, Encantado valley; 3450 m; 25°10'S, 65°52'W.
91. meg	M	500030	33	Okada 7516	Argentina. Salta: 2 h by foot from Isonza following ravine; 3500 m; 25°20′S, 65°53′W.
92. meg	M	233124	Z	CPC 2535	Bolivia. Unknown.
93. meg	M	310977	Z	Alandia 64-9	Bolivia. Unknown.
94. meg	M, D	310978	Z	Alandia 64-10	Bolivia. Unknown.
95. tor	M	310936	3	Ugent 4853	Bolivia. La Paz: Ayopaya; 10 km from Hacienda Sailapata; 16°30'S, 66°35'W.
96. tor	M, D	546014	3	Hawkes et al. 200	Bolivia. La Paz: Loayza; 11.2 km from Ayo Ayo on the road to Caracato; 3960 m; 17°03'S, 67°53'W.
97. tor	M, D	498257	4	Ochoa 11914	Bolivia. La Paz: Inquisive; Huanecta, between Tres Cruces and Quime; 4450 m; 17°04'S, 67°18'W.
98. tor	M, D	498142	5	Hawkes et al. 6572	Bolivia. Cochabamba: Ayopaya; road from Cruce de Challa to Independencia, Abra de Ornoni; 3730 m; 17°15'S, 66°54'W.

TABLE 2. Continued.

Taxon <sup>1</sup>	Study <sup>2</sup>	PI <sup>3</sup>	Area <sup>4</sup>	Collector	Locality
	Study-		Area	Collector	Locality
99. tor	M, D	498143	5	Hawkes et al. 6589	Bolivia. Cochabamba: Ayopaya; 24 km from Independencia on road to Challa; 3730 m; 17°14'S, 66°54'W.
100. tor	M, D	498144	6	Hawkes et al. 6613	Bolivia. Cochabamba: Chapare; 35 km from Tranca de Sacaba on road to Palca; 3760 m; 17°14'S, 66°03'W.
101. tor	M, D	498145	6	Hawkes et al. 6616	Bolivia. Cochabamba: Chapare; 35 km from Tranca de Sacaba; 3760 m; 17°14′S, 66°03′W.
102. tor	M	498146	6	Hawkes et al. 6622	Bolivia. Cochabamba: Chapare; 0.5 km from Palca, 40 km from Tranca de Sacaba on road to Palca; 3730 m; 17°13'S, 66°03'W.
103. tor	M, D	473389	7	Hawkes et al. 4758	Bolivia. Cochabamba: Toralapa; 71.5 km from Cochabamba on Santa Cruz road; 3500 m; 17°26'S, 65°43'W.
104. tor	M, D	545928	8	Hoopes et al. 174	Bolivia. Cochabamba: Chapare; 9 km from turn-off at Aguirre from the Cochabamba- Chapare road; 3500 m; 17°18'S, 64°44'W.
105. tor	.M, D	545892	8	Hoopes et al. 175	Bolivia. Cochabamba: Chapare; 9 km from turn-off at Aguire from the Cochabamba-Chapare road; 3500 m; 17°18'S, 64°44'W.
106. tor	M, D	545998	9	Hawkes et al. 251	Bolivia. Cochabamba: Ibañes; 18.2 km from Sacaca on road to Oruro; 3900 m; 18°00'S, 66°26'W.
107. tor	M, D	458396	10	Hawkes et al. 4556	Bolivia. Chuquisaca: Oropeza; 29 km from Su- cre toward Ravelo; 3300 m; 19°02'S, 65°17'W.
108. tor	M, D	458397	16	Hawkes et al. 4695	Bolivia. Tarija: Méndez; 32 km from Tarija to- ward Iscayachi; 3000 m; 21°31′S, 64°45′W.
109. tor	M, D	545926	17	Hoopes et al. 94	Bolivia. Tarija: Méndez; Cuesta de Sama; 2950 m; 21°29'S, 64°53'W.
110. tor	M, D	472805	19	Hoffman 1851	Argentina. Salta: 56 km E of Cajas; 3150 m; 22°15'S, 65°18'W.
111. tor	D	320303	21	Hawkes et al. 3829	Argentina. Salta: 16 to 16.7 km from Santa Victoria on road from Yavi to Santa Victo- ria; 3450 m; 22°15'S, 64°58'W.
112. tor	M	472804	21	Hoffman 1738	Argentina. Salta: 9 km SW of Santa Victoria, Cuesta Parada; 3350 m; 22°15'S, 64°58'W.
113. tor	M, D	472806	21	Hoffman 1926	Argentina. Salta: 10 km SE of Santa Victoria; 3300 m; 22°15'S, 64°58'W.
114. tor	M, D	472808	21	Okada 5436	Argentina. Salta: 50 km E of Abra Lizoite; 3500 m; 22°13'S, 65°14'W.
115. tor	M, D	472807	26	Hoffman s.n.	Argentina. Salta: Santa Victoria department; 22°25'S, 64°55'W.
116. tor	M, D	320302	33	Hawkes et al. 3727	Argentina. Salta: 21.4 km from Escoipe on road to Cachi, Cuesta del Obispo; 3200 m; 25°10'S, 65°51'W.
117. tor	M, D	195210	Z	CPC 1773	Bolivia. Unknown.
118. blv	M, D	265861	10	EBS 1847	Bolivia. Chuquisaca: 8 km from Tarabuco on road to Zudañez; 19°10'S, 64°57'W.
119. blv	M	310974	Z	Alandia 64-6	Bolivia. Unknown.
120. blv	M	310975	Z	Alandia 64-7	Bolivia. Unknown.
121. blv	M	498215	Z	Ochoa 11929	Bolivia. Chuquisaca: Ckucha Tambo, on road from Sucre to Guerraloma; 2940 m.
122. blv	D	265860	Bol	EBS 1795	Bolivia. Unknown: E of Siclla; 3600 m.

TABLE 2. Continued.

Taxon <sup>1</sup>	Study <sup>2</sup>	PI <sup>3</sup>	Area <sup>4</sup>	Collector	Locality
123. sgr	M, D	230510	Peru	Ochoa 1440	Peru. La Libertad: near Santiago de Chuco; 3550 m; 08°09'S, 78°11'W.
124. sgr	M, D	365360	Peru	Ochoa S-54	Peru. Ancash: Acrana; 3500 m; 09°30'S, 77°45'W.
125. alb	D	266381	Peru	Correll P863	Peru. Cajamarca: 6 km from the entrance to Hacienda Porcón; 3500 m.
126. pne	D	473431	Peru	Ochoa 7990	Peru. Unknown.
127. aem	D	472793	30	Okada 4361	Argentina. Jujuy: Piedra Rosada, by the Ventura River; 4000 m; 23°35'S, 65°12'W.
128. acl	D	472801	Arg	Okada 6083A	Argentina. La Rioja: Sierra de Famatina, La Encrucijada; 3050 m; 28°58'S, 67°42'W.
129. brd	D	245763	Chil	Correll C14	Chile. Region IX: road from Cherquenco to Refugio Llaima; 800 m; 38°40'S, 71°53'W.
130. blb	D	275200	Guat	Hawkes 1796	Guatemala. Huehuetenango: 5 km S of Mala- catancito, road from Huehuetenango to Quezaltenango; 1800 m.
131. can	D	265864	Peru	EBS 1831	Peru. Cuzco: 4 km on the road from Cuzco to Pisac; 3800 m.
132. rap	D	473369	Peru	Hawkes et al. 5138	Peru. Cuzco: 25 km on road from Cuzco to Puno; 3200 m.

et Buk. (2n = 24; series *Tuberosa*; Huamán et al. 1980, 1982, 1983; Johns et al. 1987). Okada and Clausen (1982) document the occurrence of natural triploid (2n = 36) hybrids between *S. meg*istacrolobum and S. acaule subsp. acaule in northwestern Argentina and formally name them S. × indunii Okada and Clausen. They further suggest that some populations of S. acaule subsp. aemulans Bitter and Wittm. (2n = 48) are hybrids between S. acaule subsp. acaule and S. megistacrolobum or S. ×indunii. They also question Hawkes and Hjerting's (1969) hypothesis of the hybrid origin of S. ×brucheri (see above) and suggest that it is related to S. gourlayi. Hawkes and Hjerting (1989) mention "intermediate forms" between S. megistacrolobum and S. toralapanum that they believe are the result of introgression between the two. They also designate natural hybrids of S. megistacrolobum with S. acaule, S. infundibuliforme, S. sparsipilum (Bitter) Juz. and Buk. (2n = 24; series Tuberosa) and S. stenotomum; and of S. toralapanum with S. acaule. Ochoa (1990) hypothesizes natural hybridization between some populations of S. megistacrolobum and S. toralapanum and S. megistacrolobum and S. infundibuliforme. He suggests a "connecting link" of *S. megistacrolobum* to series Acaulia and believes that S. megistacrolobum hybridizes naturally with S. acaule. Hawkes (1990) reverses his earlier hypothesis relative to the origin of *S.* ×*brucheri* (see above), and designates it as a hybrid between *S. gourlayi* and *S. infundibuliforme* (Hawkes and Hjerting 1969).

The objective of this study is to assess the nature, extent, and geographical partitioning of morphological differences between S. megistacrolobum, S. toralapanum, and two other species in series Megistacroloba; S. boliviense and S. sogarandinum Ochoa. We chose these latter two species from the many others mentioned above because of their close phenetic resemblance to S. megistacrolobum and S. toralapanum. A separate study (Giannattasio and Spooner 1994) examines a subset of these taxa and other species for differences in single-to low-copy nuclear DNA. These combined data will be used to assess taxonomic boundaries and possible patterns of hybridization involving S. megistacrolobum and S. toralapanum.

#### MATERIALS AND METHODS

**Plants.** This study uses 93 accessions of *S. megistacrolobum*, 22 of *S. toralapanum*, four of *S. boliviense*, and two of *S. sogarandinum* (Table 2), chosen from the germplasm bank of NRSP-6 (Hanneman and Bamberg 1986), and grown in an experimental field plot at Sturgeon Bay, Wis-

TABLE 3. Characters used in phenetic analysis of Solanum megistacrolobum and S. toralapanum. Characters with an asterisk (\*) were significantly different (0.05 level or greater) and used for a separate phenetic analysis.

Leaf characters

\*1. Leaf length (cm). \*2. Ratio: leaf width/leaf length. \*3. Length of terminal leaflet lamina (cm). \*4. Ratio: terminal leaflet width/terminal leaflet length. \*5. Ratio: distance from base of petiole to widest part of terminal leaflet/leaf length. \*6. Length of primary lateral leaflet (cm). \*7. Degree of decurrency of the primary lateral leaflet; measured by the length of the primary leaflet five mm basiscopic to the midrib (cm). 8. Density of adaxial pubescence: glabrous (0), puberulent (1), intermediate (2), dense (3). \*9. Density of abaxial pubescence: glabrous (0), puberulent (1), intermediate (2), dense (3). \*10. Parsley-like leaf odor: absent (0), slight (1), moderate (2), extreme (3). \*11. Ratio: distance from base to widest part of terminal leaflet/terminal leaflet length. \*12. Number of leaflets.

Floral characters (see Spooner and van den Berg 1992b, for illustrations of characters 22, 23, 24, and 25)

\*13. Number of flowers per inflorescence. \*14. Length of peduncle (cm). \*15. Distance from base of peduncle to articulation (cm). \*16. Position of the pedicel articulation; measured by the ratio of character 15/total pedicel length. 17. Width of calyx lobe (cm). \*18. Length of calyx lobe (cm). \*19. Ratio: length of acumen/length of calyx lobe. \*20. Posture of calyx acumen: tightly appressed (1), moderately appressed (2), spreading to recurved (3). \*21. Color of corolla: whitish blue (1), blue (2), deep blue to purple (3). \*22. Radius of corolla: longest distance between center of corolla and tip of petal (cm). 23. Petal lobe length (cm). \*24. Corolla shape; ratio between the petal lobe length and petal lobe width. 25. Ratio: radius from center of corolla to base of corolla lobe/radius of corolla. 26. Length of anther (cm).

consin. The accessions of *S. megistacrolobum* and *S. toralapanum* were chosen to represent the widest possible geographic range of these species. They are mapped into 33 generalized geographic regions (Fig. 1). The accessions were planted from seed in early June 1990; eight seedlings per accession were planted in rows in a common field plot in early July, and all measurements were taken in late August when the plants were in flower.

Character Measurement. The middle four surviving plants per row were measured for each accession. A total of 26 characters (Table 3) were measured on each plant. Of these, 21 characters were quantitative and five were qualitative. Leaf characters were measured using the fourth true leaf of each plant. Floral characters were taken from the uppermost inflorescence. The means of these four plants were used as representative of the accession. In rare cases, a character was lacking entirely (i.e., primary lateral leaflet length), and the average of that character per species was substituted for the missing character. Many herbarium specimens of members of sect. Petota collected in the wild are incomplete, have been collected at different growth stages, or have been collected under different environments. Also, many wild potato species, including S. megistacrolobum and S. toralapanum, are thought to exhibit morphological

plasticity under different environments (Brücher 1959; Correll 1962; Hawkes 1990; Hawkes and Hjerting 1989; Ochoa 1990). Our study with living plantings grown in one location, with measurements taken from serial organs at the same growth stage, standardizes environments and measurements of organs at the same serial stage.

Data Analysis. Each character was analyzed for its mean, range, standard deviation, and significance by one-way ANOVA in Minitab (Ryan et al. 1985). These analyses were performed twice, once for all taxa, and again for a subset of the taxa that occurred in the 33 geographic areas that had only one species per area (i.e., omitting all accessions from areas 4, 7, 16, 19, 21, 26, and 33). This second analysis was done to test whether morphological intermediacy between species was caused by hybridization or primary divergence. Phenetic analyses of all accessions were produced by NTSYS-pc®, version 1.70 (Rohlf 1992). Averaged data for each character were standardized (STAND) and similarity matrices [in SIMINT, using average taxonomic distance (DIST), Euclidean distance (EUCLID), Manhattan distance (MANHAT), and product-moment correlation (CORR)] were generated. Clustering was performed using the unweighted pair-group method (UPGMA) in SAHN. Cophenetic correlation coefficients (COPH, in MXCOMP) were used to measure distortion between the similarity matrices and the resultant four phenograms (Rohlf and Sokal 1981; Sokal 1986). These analyses were performed twice, once for all characters, and again for a subset of the characters that were significantly different (0.05 level) between *S. megistacrolobum* and *S. toralapanum*. Principal Components Analyses (PCA) also were performed from similarity matrices (CORR) and EIGEN. As in the UPGMA analysis, PCA were performed twice, once for all characters, and again for a subset of the characters that were significantly different (0.05 level) between *S. megistacrolobum* and *S. toralapanum*.

#### RESULTS

One-way ANOVA of the entire data set demonstrated that 21 of the 26 characters (81%), including the four major characters used previously to distinguish S. megistacrolobum and S. toralapanum (Table 1), were significantly different (0.05 level or greater) between them. The five characters not showing this significance were characters 8, 17, 23, 25 and 26 (Table 3). Our data show extensive intra-accession, intraspecific and interspecific variability for most characters (Fig. 2). For example, while none of the S. toralapanum examined were found to possess the parsley-like leaf odor characteristic of S. megistacrolobum, 74 out of the 93 accessions of S. megistacrolobum had at least one of the four individuals per accession lacking this odor. Many of these accessions also had individuals with strong odor, however, demonstrating high intra-accession variability for this character.

One-way ANOVA of the reduced data set that eliminated taxa from areas of general geographic co-occurrence of *S. megistacrolobum* and *S. toralapanum* (see above) demonstrated that 19 of the 26 characters (73%), including the four major characters used previously to distinguish them, were significantly different (0.05 level or greater). In general, however, there was little difference in levels of significance of characters relative to the entire data set. All further analyses used all accessions.

The phenograms using all 26 characters generated by DIST and EUCLID had the greatest cophenetic correlation coefficients (0.87), were equal in topology, and differed only in the scaling of the phenograms. These phenograms,

however (not shown here, but see Giannattasio 1992), intermixed 17 accessions of *S. megistacrolobum* and 21 accessions of *S. toralapanum* on one cluster of the phenogram, and therefore, provided poor separation of the two species.

The phenograms using only the 21 statistically significant characters did a much better job of distinguishing *S. megistacrolobum* from *S. toralapanum*. As above, the phenograms generated by DIST and EUCLID had the greatest cophenetic correlation coefficients (0.83), were equal in topology, and differed only in the scaling of the phenograms. The phenogram generated by DIST is shown in Figure 3. The cophenetic correlation coefficients for the phenograms generated by CORR and MANHAT were 0.65 and 0.77, respectively.

This second analysis, using only those characters showing statistically significant differences between S. megistacrolobum and S. toralapanum, largely separated them into two separate clusters of the phenogram, with the exception of five of the 93 accessions of S. megistacrolobum that clustered with S. toralapanum, and two that clustered between S. boliviense and S. sogarandinum. Also, one accession of S. toralapanum clustered with three of the four accessions of S. boliviense. Both accessions of S. sogarandinum and three of the four accessions of S. boliviense clustered among themselves, separated by the two "misplaced" accessions of S. megistacrolobum. The remaining accession of S. boliviense clustered with two of the five "misplaced" accessions of S. megistacrolobum in the S. toralapanum cluster (Fig. 3).

PCA using all 26 characters (not shown here, but see Giannattasio 1992), like the UPGMA, provided somewhat poor separation of *S. megistacrolobum* and *S. toralapanum* by showing contiguous distribution of taxa along the first three PCA axes, with much interdigitation between species. The first three principal components axes accounted for 24.1%, 12.5%, and 7.8%, for a total of 44.4% of the total variation.

PCA of the reduced data set using the 21 statistically significant characters also showed contiguous distribution but with less interdigitation of taxa (Fig. 4). This analysis, like the UPGMA analysis, grouped both accessions of *S. sogarandinum*, but provided a better grouping of all four accessions of *S. boliviense*. The first three principal components axes accounted for 27.7%, 13.9%, and 9.8%, for a total of 51.4% of the total variation. The six highest loadings on PCA axis

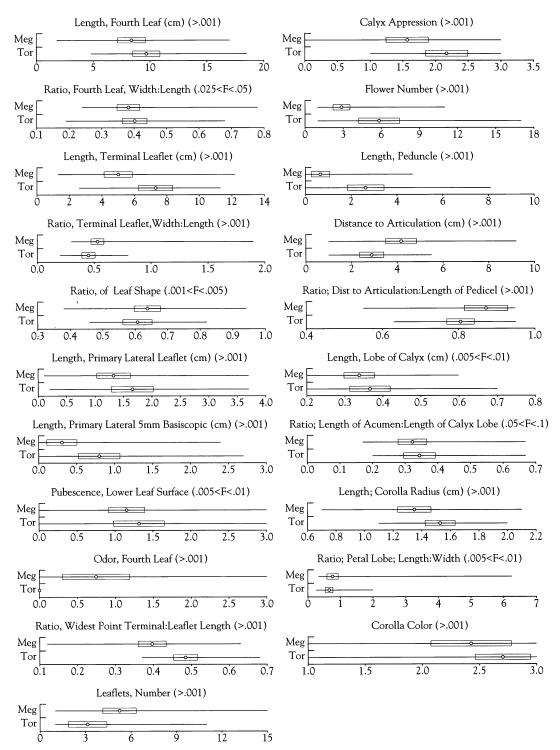


Fig. 2. Means, ranges and one standard deviation of the mean for the 21 characters showing significant differences (0.05 level or greater) between *Solanum megistacrolobum* (meg) and *S. toralapanum* (tor). See Table 3 for an explanation of character states.

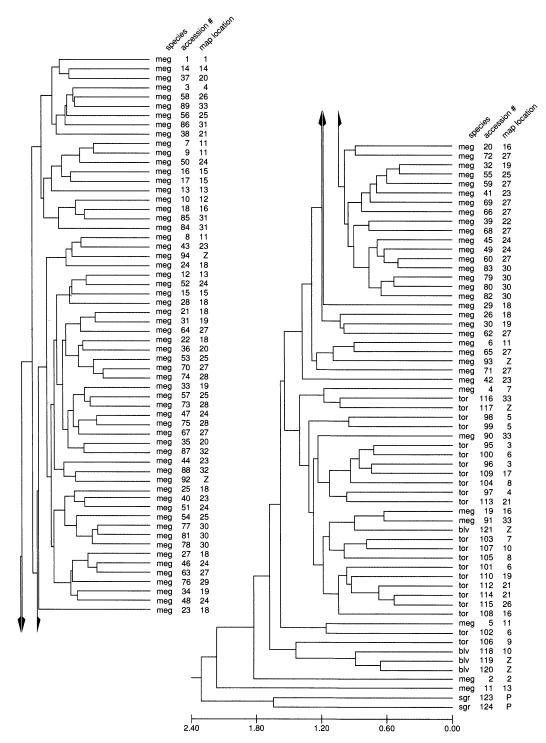
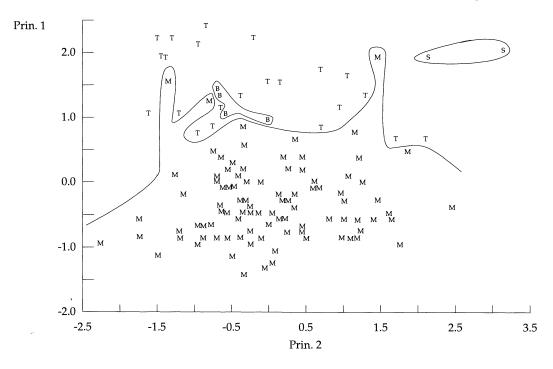


FIG. 3. UPGMA phenogram (DIST similarity option) based on the 21 of the 26 morphological characters that showed statistically significant differences between Solanum megistacrolobum (meg) and S. toralapanum (tor). Solanum boliviense = blv; S. sogarandinum = sgr. See Table 2 for a listing of accession numbers and map locations. The letters refer to locations outside of Figure 1. P = Peru; Z = unknown location.



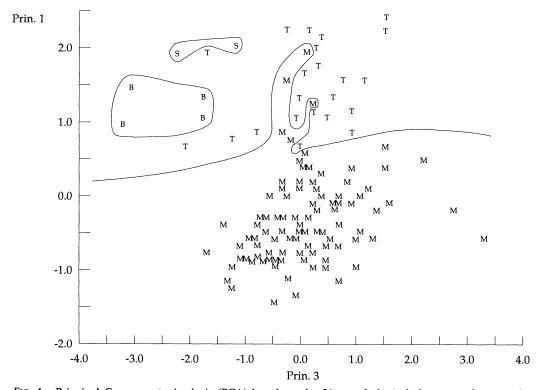


Fig. 4. Principal Components Analysis (PCA) based on the 21 morphological characters shown to be significantly different (0.05 level or greater) between Solanum megistacrolobum and S. toralapanum. B = Solanum boliviense, M = S. megistacrolobum, S = S. sogarandinum, and T = S. toralapanum.

one were for characters 14, 3, 13, 22, 11, and 10; the six highest loadings on PCA axis two were for characters 5, 6, 12, 21, 4, and 7 (Table 3). Three of these characters (7, 10, and 14) have been used by prior taxonomists to distinguish *S. megistacrolobum* and *S. toralapanum* (Table 1).

#### DISCUSSION

Prior descriptions of *S. megistacrolobum* and *S.* toralapanum largely distinguished the taxa by four morphological characters. These characters are so highly variable (Fig. 2), often within individual accessions, that reliance on any one of them to identify taxa could lead to frequent misidentifications. The accessions we analyzed, however, largely clustered by prior determinations, but only after eliminating the five characters that showed no statistically significant differences between S. megistacrolobum and S. toralapanum. This indicates that the morphological characters are useful, even though individual character states regularly cross species boundaries. These accessions have been identified by various taxonomists at NRSP-6 from field plantings of from 6 to 12 plants per accession, and have been accompanied by data on prior determinations and locality data (Spooner and van den Berg 1992a). It is apparent that reliably consistent determinations for some accessions require the use of many characters from more than one individual per accession. We assume that the patterns of intra-and interspecific variability documented here would be similar in replicated field plantings over different environments, but our conclusions are based on this one field planting. It also is possible that these taxa would better maintain their differences in their native environments.

The high degree of variability within these two species, and intermediacy between them, is consistent with prior observations (Brücher 1959; Correll 1962; Hawkes 1990; Hawkes and Hjerting 1989; Ochoa 1990). The high morphological variability we documented within *S. megistacrolobum* also is consistent with the high variability in their glycoalkaloid profiles shown by Johns and Osman (1986). They interpret their data to group *S. toralapanum* with *S. boliviense* (and *S. sanctae-rosae*). Our morphological data showed that most accessions of *S. megistacrolobum* and *S. toralapanum* are more similar to each other than either is to *S. boliviense*.

Some of the intermediacy between S. megis-

tacrolobum and S. toralapanum is partitioned in zones of geographical overlap between their main areas of distribution. Four of the five accessions of S. megistacrolobum that cluster with S. toralapanum (Fig. 3) co-occur with this species in areas 7, 16, and 33 (Table 2; Fig. 1). This pattern is consistent with hypotheses of interspecific hybridization, primary divergence, or misidentification. The remaining three "misplaced" accessions of S. megistacrolobum are from areas 2, 11, and 13 where only one species per area occurs. The one-way ANOVA of character state differences between species for the subset of the taxa that occur in the geographic areas that had only one species per area show a similar to slightly reduced significance of characters relative to the entire data set. These results would be expected when hybridization or primary divergence were not occurring. The distributional data, therefore, are ambiguous regarding causes of morphological intermediacy between S. megistacrolobum and S. toralapanum.

Our study demonstrates the following between S. megistacrolobum and S. toralapanum: 1) there is much intraspecific variation in all character states, 2) there are intermediate populations that appear to bridge the gap between the species, 3) no morphological characters can be used alone to distinguish the taxa consistently, 4) the taxa have contiguous and somewhat overlapping distributions, 5) the taxa are difficult to differentiate consistently, especially on the basis of key characters using single specimens, and 6) the taxa can generally be distinguished, but only by multivariate techniques, and then only with a subset of the characters. Patterns of overlapping character states used to define similar taxa have been shown to be common throughout much of Solanum sect. Petota (Spooner and van den Berg 1992a). In some cases, these taxa have not been supported by morphological reinvestigations (Spooner et al. 1993; van den Berg and Spooner 1992). Other studies, however (Spooner and van den Berg 1992b), and this one, provide weak support for the recognition of taxa at some level. The similarity of these taxa, and resulting confusion as to taxonomic status among similar taxa is reflected in widely different treatments of sect. Petota by different contemporary authors (Spooner and van den Berg 1992a). The differing contemporary taxonomic treatments of S. megistacrolobum and S. toralapanum as species (Gorbatenko 1989; Hawkes and Hjerting 1989;

Hawkes 1990) or as varieties (Ochoa 1990) is one example of such disagreement.

Our experience, based on the results of the one-way ANOVA of character states, loadings on the PCA, and our personal field observations suggest that the following combination of characters best differentiate *S. megistacrolobum* and *S. toralapanum*: leaf odor, flower number, decurrency of the primary lateral leaflets, and peduncle length. The wide variability of these characters makes the construction of a dichotomous key difficult, and we refer the reader to Figure 2 for useful means and ranges of these character states.

The morphological and distributional patterns of S. megistacrolobum and S. toralapanum, that of weakly differentiated taxa, with contiguous to slightly overlapping geographic distributions, more closely fits a model of subspecies or varietal rank than one of separate species (Stuessy 1990). One biological factor also often associated with varieties or subspecies is reduced crossability between them, but this reduced crossability is not known to occur between these two taxa (Buck 1966; Hawkes 1990; Ochoa 1990; Pandey 1960), including hybrids to the F<sub>2</sub> generation (Hawkes and Hjerting 1989). Solanum sect. Petota includes many phenetically very distinct taxa, some with different geographic ranges, that have the ability to produce advanced generation hybrids artificially, with little or no reduction in fertility (Spooner and van den Berg 1992b). Hawkes (1990) hypothesizes that many wild potato taxa are reproductively isolated by cryptic structural differences of the chromosomes and that artificial and natural hybrids exhibit advanced generation breakdown. His data, however, are largely observational and lack detailed measurements, intraspecific crossing controls, and statistical presentation of results, and are therefore questionable (Spooner and van den Berg 1992a). A strict application of the biological species concept in much of sect. Petota, therefore, would lump many taxa that have distinct morphological (and perhaps agronomically important) characters. Therefore, we rely on a morphological species concept regarding S. megistacrolobum and S. toralapanum. For these reasons, our data agree most closely with Ochoa's (1990) treatment of S. toralapanum as a variety of S. megistacrolobum [as S. megistacrolobum var. toralapanum (Cárd. and Hawkes) Ochoa]. Further discussion of taxonomic rank is provided in Giannattasio and Spooner (1994).

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